

# The frequency of CMV and EBV infections before liver transplantation in patients on the waiting list for liver transplantation referring to a liver transplant center, the North of Iran

Reihaneh Sadeghi Garmaroodi<sup>1</sup>, Hamed Naziri<sup>2</sup>, Pirooz Samidoust<sup>3,\*</sup>

<sup>1</sup>Razi Clinical Research Development Unit, Razi Hospital, Guilan University of Medical Sciences, Rasht, Iran

<sup>2</sup>Department of Microbiology, School of Medicine, Guilan University of Medical Sciences, Rasht, Iran

<sup>3</sup>Department of Surgery, School of Medicine, Guilan University of Medical Sciences, Rasht, Iran

## Abstract

Viral infections, such as cytomegalovirus (CMV) and Epstein-Barr virus (EBV), contribute to the low survival rates of liver transplant patients. This study aimed to assess the prevalence of CMV and EBV infections in patients awaiting liver transplantation. Utilizing a census sampling approach, this cross-sectional study examined all cases of viral infections from 2016 to 2021 among liver transplant patients referred to Rasht liver transplant center, the North of Iran. In total, 34 individuals with a mean age of  $48.9 \pm 12.2$  years were included in the study. Of these, 40 individuals (59.7%) were male. The prevalence of CMV IgM and IgG antibodies among liver transplant candidates was 7.5%, and 97%, respectively. Also, the prevalence of EBV IgM and IgG antibodies was 7.5%, and 97%, respectively. The average serum vitamin D level in CMV IgM-negative patients was  $30.7 \pm 17.2$  compared to  $55.1 \pm 22.1$  in CMV IgM-positive patients ( $p = 0.011$ ). The prevalence of CMV and EBV infections in liver transplant patients was found to be 7.5%. These results highlight the necessity for continuous and effective strategies to prevent infection-related complications through prompt diagnosis and treatment, which are crucial for positive liver transplant outcomes.

**Keywords:** Cytomegalovirus, Epstein-Barr virus, Liver transplantation, Transplant recipient, Transplant donor

## 1. Introduction

Liver transplant recipients face numerous challenges, both immediate and long-term post-transplantation. Short-term issues can be categorized into technical complications, such as venous thrombosis and biliary disorders, and medical concerns, including infections and acute transplant rejection [1]. Post-liver-transplant infections have emerged as a primary cause of morbidity and mortality, exceeding acute rejection

rates, despite advancements in surgical techniques and immunosuppressive therapies [2, 3].

Post-transplant infections are classified as either early or late, with late infections predominantly resulting from immunosuppressive drug effects, whereas early infections have alternative etiologies [3]. Enhancing our understanding of prevalent post-transplant infections and their risk factors will facilitate the development of strategies to mitigate these risks and prevent subsequent infections.

### \*Corresponding author:

Pirooz Samidoust, MD  
Razi Hospital, Guilan University of Medical Sciences,  
Rasht, Iran  
Tel/Fax: +98 13 33542460  
Email: piroozesamidoost@gmail.com  
<http://orcid.org/0000-0002-7916-0677>

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Multiple independent risk factors contribute to the development of bacterial, fungal, and viral infections following transplantation. Multiple risk factors for increased post-transplant infection risk have been identified in studies. The majority of studies found prolonged operation time, extended hospitalization post-transplant, and elevated Model for End-Stage Liver Disease (MELD) scores as significant predictors of infection risk [4-6]. The MELD scoring system is favored over the Child-Turcotte-Pugh (CTP) system for patient classification and outcome prediction post-liver transplantation [7-9]. Additionally, intraoperative transfusions, particularly of Fresh Frozen Plasma (FFP) and packed red blood cells, are recognized as post-liver-transplant infection risk factors [10, 11].

Cytomegalovirus (CMV), a member of the herpesvirus family, is a significant pathogen in organ transplant recipients, affecting approximately 50-60% of this population [12-15]. It is the most significant virus that can infect humans and can cause a range of severe clinical syndromes, including fever, leukopenia (mononucleosis-like syndrome), hepatitis, pneumonitis, pancreatitis, colitis, meningoencephalitis, and gastrointestinal bleeding in immunocompromised individuals, including organ transplant recipients [16]. Around 6% of healthy adults are asymptomatic carriers of CMV [17], and nearly two-thirds of transplant recipients have pre-existing immunity, indicated by serum anti-CMV IgG antibodies [18]. The virus is transmissible via sexual or respiratory routes, blood transfusions, and from mother to child during birth or breastfeeding. Transplant recipients with positive anti-CMV IgG antibodies are at risk for latent virus reactivation, typically between 1-4 months post-transplantation [16,18]. This reactivation is influenced by the preoperative serostatus of both the recipient and donor, as well as the type and dosage of immunosuppressive medications used [19]. The administration of anti-T cell antibodies, high-dose corticosteroids, and Mycophenolate mofetil have been associated with increased disease severity [20]. Some clinical studies have also suggested a link between CMV infection and graft rejection, with Schnitzler et al. reporting that mortality rates were 2.5 times higher in patients with pre-transplant CMV antibodies who did not receive ganciclovir prophylaxis compared to those who did [21].

Another contributor to these infections is the Epstein-Barr virus (EBV). EBV, a ubiquitous human virus found worldwide, typically spreads through bodily fluids, especially saliva, and can cause infectious mononucleosis. EBV infection can exacerbate immune system suppression, especially when combined with immunosuppressive drug therapy. Some researchers posit that EBV infection may disrupt and stimulate the immune system, increasing the risk of transplant rejection [4]. It is estimated that 80 to 90 percent of transplant recipients develop a secondary EBV infection within the first year following transplantation, which closely correlates with graft dysfunction [5]. EBV reactivation can provoke an immune response, potentially leading to transplant rejection. In some cases, increased immunosuppression to counteract rejection may result in patient mortality, highlighting the clinical significance of monitoring EBV infection [22].

Research conducted at the Pasteur Institute of Iran virology department, as well as a study on EBV infection in Tehran children and adults, suggest that approximately 80% of transplant recipients are affected by EBV [23]. Kenagy's study in the United States reported an incidence of secondary EBV infection of 17.4% within one year following transplantation. The study results indicated that 33% of the patients exhibited positive serology [24]. However, Acott's Canadian research observed EBV reactivation in 12.5% of patients experiencing acute graft rejection [25]. Shahinian et al. indicated that primary EBV infection might play a pathogenic role in some cases of post-transplant lymphoproliferative disorder (PTLD) [26].

Given the growing reliance on liver transplantation as a treatment for liver disease, meticulous attention to transplant candidates' care is imperative both pre- and post-transplantation. Viral infections, notably CMV and EBV, are recognized as factors that diminish liver transplant survival rates. Therefore, thorough serological and molecular screening for these viruses is vital. Comprehending the influence of viral infections on the survival of liver transplants can enhance patient longevity and transplant success rates, and these insights can be useful for the management of liver transplant recipients, particularly in the Guilan province.

## 2. Materials and Methods

### 2.1 study design

This cross-sectional analytical study was conducted at the liver transplant center of Razi Hospital in Guilan province. The study population comprised liver transplant patients who presented to the center between 2016 and 2021. A total of 120 liver transplant candidates or recipients were evaluated for viral infections. Data collection was performed using a census approach, encompassing all relevant information from approximately 120 liver transplant patients at Razi Hospital, the North of Iran. Patient data, including age, gender, occupation, risk factors, underlying diseases, time elapsed since transplantation, and post-transplant medications, were extracted from hospital records. The analysis included only patients with complete data sets, excluding those with incomplete records. Laboratory results of CMV and EBV viral infections were obtained and included in the study.

### 2.2 Data Analysis

Descriptive statistical methods, including mean, standard deviation, frequency, and percentage, were employed to characterize the descriptive data. The independent samples t-test was utilized for the analysis of quantitative variables. These analyses were conducted using IBM SPSS software version 28, with a significance level set at  $P < 0.05$  for all tests.

## 3. Results

The study evaluated 34 individuals with a mean age of  $48.9 \pm 12.2$  years. Of these participants, 40 (59.7%) were male, and the rest were female. The mean duration post-transplantation was  $2.5 \pm 1.8$  years, ranging from 0.2 to 6 years.

The frequency of CMV infection, with CMV IgM and IgG positivity rates among liver transplant candidates being 7.5% ( $n=5$ ) and 97% ( $n=65$ ), respectively. A dual positivity for IgM in both CMV and EBV was observed in only two (3%) patients.

The study found that five (7.5%) and 65 (97%) liver transplant candidates were positive for EBV IgM and IgG, respectively.

The mean serum vitamin D level was  $30.7 \pm 17.2$  in CMV IgM-negative patients and  $55.1 \pm 22.1$  in CMV IgM-positive patients, with a significant difference observed ( $P = 0.011$ ). The average serum vitamin D level was  $31.8 \pm 18.9$  in EBV IgM-negative patients

and  $43.8 \pm 13.9$  in EBV IgM-positive patients, with no significant difference ( $P = 0.224$ ). Table 5 compares the serum vitamin D levels in patients with who co-infected with CMV and EBV infections who tested positive for IgM. No significant difference was found in serum vitamin D levels in IgM-positive individuals who infected with CMV and EBV simultaneously ( $p = 0.372$ ).

## 4. Discussion

Given the growing prevalence of liver transplantation as a therapeutic option for patients with liver disease, meticulous care for transplant candidates both pre- and post-operation is imperative. Literature reviews suggest that viral infections, such as those caused by CMV and EBV, contribute to reduced post-transplant survival rates. Exploring the impact of these viral infections on the success of liver transplants may enhance both the longevity of the transplant and patient survival. The results indicated that five (7.5%) of the liver transplant candidates tested positive for CMV IgM and 65 (97%) for IgG. Jamalidoust et al. 2021 retrospective study at Namazi Hospital in Shiraz, which sought to quantify CMV load and assess clinical outcomes in liver recipients with reactivated CMV infection, included 657 patients who received transplants from 2014 to 2017. Diagnoses were made using the real-time PCR method. The study found that 151 patients (23%) experienced CMV reactivation at least one-year post-transplant. Of these, 41 individuals (6.2%) died, and 58 (8.8%) faced transplant rejection within the first year following their surgery. Among the deceased, 21 had experienced CMV reactivation. The mortality rate was notably higher in patients with CMV infections compared to those without [27]. Conversely, our study revealed that 97% of patients are at risk of reinfection due to IgG positivity.

In a 2020 retrospective cohort study, Fernandes Garcia et al. examined the incidence of CMV disease within the first six months post-transplant among liver transplant recipients in Mexico City. Out of 124 patients, four (3.2%) contracted CMV, 96 (85%) exhibited detectable DNAemia, and 25 (22%) remained asymptomatic. The study concluded that the incidence of CMV disease was 3.2% [28], a relatively minor proportion compared to the IgG-positive individuals in our study who may be

susceptible to future CMV infections. Moreover, in 2017, Varghese et al. investigated CMV seropositivity in liver transplant recipients. The pre-transplant analysis focused on CMV-related IgG and IgM. Overall, CMV exposure in recipients was found to be 71.8%. Among donors, CMV seropositivity was observed in 90.9% (100 out of 110). Notably, three deaths occurred in recipients who were also positive for CMV via quantitative RT-PCR. The findings of the study indicate a high rate of CMV exposure among both transplant recipients and donors, with the greatest risk associated with recipient reactivation rates. However, the mortality rate due to CMV reactivation was low [29].

In 2013, Dehghani et al. assessed the prevalence of CMV serology in pediatric liver transplant candidates at Namazi Hospital in Shiraz. This descriptive, cross-sectional, and retrospective study analyzed serology data from 98 liver transplant candidates under 18 years old, who were referred to Namazi Hospital between 2006 and 2009. Serological testing for IgM and CMV IgG was conducted using the ELISA method. The research revealed that 92.9% of the pediatric candidates tested positive for IgM and 17.3% for CMV IgG, while 7.1% and 82.7% tested negative for IgM and CMV IgG, respectively, indicating a higher exposure rate compared to our study's subjects [30].

All volunteers and donors must undergo CMV-IgG serology testing prior to transplantation. Recipients lacking CMV antibodies face the highest risk of infection when receiving organs from antibody-positive donors, with rates up to 88% in the absence of prophylaxis. Conversely, the risk is lowest for recipients with negative antibodies receiving organs from similarly negative donors [31]. In light of these findings, Shahinian et al. recommend the use of appropriate prophylactic medications, cytomegalovirus vaccination, and vigilant patient monitoring for viral infections to mitigate CMV infection risks in transplant recipients [26]. Gane et al. conducted a study to evaluate the safety and efficacy of oral ganciclovir in preventing CMV disease following liver transplantation. In this research, 304 liver transplant recipients were randomized to receive either oral ganciclovir at a dosage of 1000 mg or a matching placebo, administered three times daily. The medication was continued until the 98th day post-transplantation, provided the patient could

tolerate oral intake. In the initial six months following surgery, patients underwent regular monitoring for indications of CMV infection, CMV disease, graft rejection, opportunistic infections, and potential drug toxicity. The study's findings revealed that the six-month incidence of CMV disease, as estimated by Kaplan-Meier, was 29 cases (18.9%) in the placebo group of 154 patients, compared to seven cases (4.8%) in the ganciclovir group of 150 patients ( $p < 0.001$ ). Among the high-risk seronegative recipients (R-) receiving seropositive livers (D+), the incidence of CMV disease was 11 cases (44%) out of 25 in the placebo group, versus three cases (14.8%) out of 21 in the ganciclovir group ( $p = 0.02$ ). A significant reduction in CMV disease incidence was observed in antibody recipients, with 12 cases (32.9%) out of 37 in the placebo group and only two cases (4.6%) out of 44 in the ganciclovir group ( $p = 0.002$ ). Oral ganciclovir also decreased the incidence of CMV infection (placebo group: 79 cases (51.5%) out of 154; ganciclovir group: 37 cases (24.5%) out of 150; ( $p < 0.001$ )) and symptomatic herpes simplex infections (Kaplan-Meier estimates: placebo group: 36 cases (23.5%) out of 154; ganciclovir group: five cases (3.5%) out of 150; ( $p < 0.001$ )). Overall, the researchers concluded that oral ganciclovir is a safe and effective prophylactic for CMV disease post-liver transplantation [32]. It is important to note that serological methods for diagnosing CMV infection can lead to delays that impact patient follow-up. Prompt and timely diagnosis of CMV, which serological tests, such as ELISA cannot provide, is crucial for transplant patient management. In such instances, antigen testing may facilitate the timely detection of the virus.

The study results revealed that five (7.5%) of liver transplant candidates tested positive for EBV IgM and 65 (97%) for IgG. In Abdullatif et al. study in London, which included 96 pediatric liver transplant patients, the incidence of EBV was found to be 60.4% [33], a notably high rate. Varghese et al., in 2017, examined the seropositivity of the Epstein-Barr viral capsid antigen (EBVCA) in liver transplant donors. Analysis of pre-transplant data from 153 recipients showed that 61.4% [29] had antibodies against EBVCA, mirroring the high rates observed in our study. Halliday et al. 2014 retrospective study assessed the prevalence of EBV in the blood and clinical outcomes of 98 liver transplant recipients.

Monitoring EBV DNA levels via whole blood PCR correlated with clinical parameters over a median period of 249 days, revealing that 67% of patients had the EBV blood virus [34], indicating a substantial prevalence of EBV infection among liver transplant patients.

Research conducted at the Pasteur Institute of Iran virology department, as well as a study on EBV infection in Tehran children and adults, suggest that approximately 80% of transplant recipients are affected by EBV [23]. Kenagy's American study reported a 17.4% rate of secondary infection within a year post-transplant [24], while Acott's Canadian research found a 12.5% reactivation rate of EBV among patients experiencing acute graft rejection (25). Rostamzadeh et al. 2007 study, consistent with other countries' findings, indicated viral reactivation from a latent to an active secondary state post-transplant. The study investigated potential causes of secondary EBV reactivation, including the use of ALG medication, acute transplant rejection, and immunosuppressive drugs; however, none of these factors were determined to be significant in virus reactivation [35].

The results of our study indicated that the average serum level of vitamin D in CMV IgM-negative individuals was  $30.72 \pm 17.17$ , while it was  $55.12 \pm 22.07$  in CMV IgM-positive individuals. For CMV IgG, the average serum levels were  $44.35 \pm 54.94$  in negatives and  $32.36 \pm 17$  in positives. Notably, a significant difference was observed in the mean serum level of vitamin D in CMV IgM infections ( $p=0.011$ ), with higher levels in CMV IgM-positive patients. Contrary to the trend in most studies, this study found no significant difference in the mean serum level of vitamin D in EBV IgM infections. The question arises as to why, against the backdrop of existing literature, vitamin D levels are elevated in CMV IgM-positive patients [36]. Generally, research suggests that vitamin D does not significantly impede CMV proliferation in vitro. Instead, CMV replication swiftly downregulates the expression of the vitamin D receptor gene, a phenomenon specifically associated with CMV and not typically seen in other viral infections, including EBV. Disruptions in vitamin D homeostasis may influence over 80 pathways linked to cancer, autoimmune diseases, and cardiovascular conditions, potentially predisposing CMV patients to

a range of disorders [36]. Bearde et al. study identified a correlation between lower calcitriol levels and heightened perinatal and early postnatal CMV transmission. However, it remains unclear whether CMV and/or HIV infections diminish vitamin D levels due to the body's increased utilization in combating these infections, or if low maternal vitamin D contributes to a higher susceptibility to viral infections [37].

The findings of this study indicate no significant differences in CMV and EBV infection rates when considering gender, liver enzymes, serum calcium, and serum albumin levels. Our research highlights that the correlation between background variables and CMV/EBV infections yields inconsistent results across various studies. Shirafkan et al. (2016) explored demographic factors and risk factors influencing the onset of cytomegalovirus (CMV) infection post-kidney transplantation. Their findings showed no correlation between the onset of CMV infection and variables such as gender, residence, marital status, education level, BMI, smoking status, hepatitis B, or dialysis type. The only variable associated with the onset of CMV infection was the patient's age, with older patients being more susceptible to CMV infection post-transplantation. Consequently, it is advisable to conduct more frequent follow-ups during the first four months post-transplantation, particularly within the initial two months [38]. Halliday et al. assessed the prevalence of the EBV blood virus and its clinical outcomes in 98 liver transplant recipients through a retrospective study, noting a prolonged infection duration in male patients [34].

This study has limitations, notably its focus on a patient group in Rasht city, which may not reflect the broader Iranian population. Therefore, future research with larger sample sizes across different cities and provinces is recommended. Systematic reviews and meta-analyses are also among our suggestions, which could also synthesize findings from various studies effectively. Further similar research is also encouraged to examine the role of vitamin D in preventing CMV infection in liver transplant recipients more closely.

Our study's results demonstrate a 7.5% incidence of CMV and EBV infections among liver transplant patients. These findings underscore the necessity of a sustained and targeted program to

avert infection-related complications through prompt diagnosis and treatment, which are crucial for favorable outcomes in liver transplant patients.

### Authors' contributions

All authors contributed equally in all parts of manuscript, also read and approved the final version of manuscript.

### Conflict of interests

None to declare.

### Ethical declarations

The Research Ethics Committee of Guilan University of Medical Sciences has been accepted all processes of the current study (IR.GUMS.REC.1401.310).

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## References

1. Moreno R, Berenguer M. Post-liver transplantation medical complications. *Ann Hepatol.* 2006;5(2):77–85.
2. Dharnidharka VR, Stablein DM, Harmon WE. Post-Transplant Infections Now Exceed Acute Rejection as Cause for Hospitalization: A Report of the NAPRTCS 1. *Am J Transplant.* 2004;4(3):384–9.
3. Bowden RA, Ljungman P, Snyderman DR. *Transplant infections.* Lippincott Williams & Wilkins; 2012.
4. Murray KF, Carithers Jr RL. AASLD practice guidelines: evaluation of the patient for liver transplantation. *Hepatology.* 2005;41(6):1407–32.
5. Freeman Jr RB, Wiesner RH, Harper A, McDiarmid S V, Lake J, Edwards E, et al. The new liver allocation system: moving toward evidence-based transplantation policy. *Liver Transplant.* 2002;8(9):851–8.
6. Lucey MR, Brown KA, Everson GT, Fung JJ, Gish R, Keeffe EB, et al. Minimal criteria for placement of adults on the liver transplant waiting list: a report of a national conference organized by the American Society of Transplant Physicians and the American Association for the Study of Liver Diseases. *Liver Transplant Surg.* 1997;3(6):628–37.
7. Wiesner RH, McDiarmid S V, Kamath PS, Edwards EB, Malinchoc M, Kremers WK, et al. MELD and PELD: application of survival models to liver allocation. *Liver Transplant.* 2001;7(7):567–80.
8. Cholongitas E, Papatheodoridis G V, Vangeli M, Terreni N, Patch D, Burroughs AK. Systematic review: the model for end-stage liver disease—should it replace Child-Pugh's classification for assessing prognosis in cirrhosis? *Aliment Pharmacol Ther.* 2005;22(11-12):1079–89.
9. Rana A, Hardy MA, Halazun KJ, Woodland DC, Ratner LE, Samstein B, et al. Survival outcomes following liver transplantation (SOFT) score: a novel method to predict patient

- survival following liver transplantation. *Am J Transplant.* 2008;8(12):2537–46.
10. Alexander J, Limaye AP, Ko CW, Bronner MP, Kowdley K V. Association of hepatic iron overload with invasive fungal infection in liver transplant recipients. *Liver Transplant.* 2006;12(12):1799–804.
11. Chow JK, Werner BG, Ruthazer R, Snyderman DR. Increased serum iron levels and infectious complications after liver transplantation. *Clin Infect Dis.* 2010;51(3):e16–23.
12. Rubin RH. Cytomegalovirus in solid organ transplantation. *Transpl Infect Dis.* 2001;3(4):1–5.
13. Falagas ME, Snyderman DR. Recurrent cytomegalovirus disease in solid-organ transplant recipients. In: *Transplantation proceedings.* 1995. p. 34–7.
14. Fishman JA, Rubin RH. Infection in organ-transplant recipients. *N Engl J Med.* 1998;338(24):1741–51.
15. Tolk O RR. Viral infections in organ transplantation. 1998;10(1):1060–3.
16. Mandell B, Bennett JE, R D Mandell B. *Dolin: Principles and Practice of Infectious Diseases.* Churchill Livingstone An Impr Elsevier Copyr 2005. 2005;4(2):1864–90.
17. Borchers AT, Perez R, Kaysen G, Ansari AA, Gershwin ME. Role of cytomegalovirus infection in allograft rejection: a review of possible mechanisms. *Transpl Immunol.* 1999;7(2):75–82.
18. Schroeder R, Michelon T, Fagundes I, Bortolotto A, Lammerhirt E, Oliveira J, et al. Cytomegalovirus disease latent and active infection rates during the first trimester after kidney transplantation. In: *Transplantation proceedings.* Elsevier; 2004. p. 896–8.
19. Sagedal S, Nordal KP, Hartmann A, Sund S, Scott H, Degre M, et al. The impact of cytomegalovirus infection and disease on rejection episodes in renal allograft recipients. *Am J Transplant.* 2002;2(9):850–6.
20. ter Meulen CG, Wetzels JFM, Hilbrands LB. The influence of mycophenolate mofetil on the incidence and severity of primary cytomegalovirus infections and disease after renal transplantation. *Nephrol Dial Transplant.* 2000;15(5):711–4.
21. Schnitzler MA, Lowell JA, Hmiel SP, Hardinger KL, Liapis H, Ceriotti CS, et al. Cytomegalovirus disease after prophylaxis with oral ganciclovir in renal transplantation: the importance of HLA-DR matching. *J Am Soc Nephrol.* 2003;14(3):780–5.
22. Rostaing L, Wéclawiak H, Mengelle C, Kamar N. Viral infections after kidney transplantation. *Minerva Urol e Nefrol Ital J Urol Nephrol.* 2011;63(1):59–71.
23. Khorvash F. Cytomegalovirus Infection in Renal Recipients in Al Zahra Hospital of Isfahan. *J Jahrom Univ Med Sci.* 2007;5(3):15–21.
24. Kenagy DN, Schlesinger Y, Weck K, Ritter JH, Gaudreault-Keener MM, Storch GA. Epstein-Barr virus DNA in peripheral blood leukocytes of patients with posttransplant lymphoproliferative disease. *Transplantation.* 1995;60(6):547–54.
25. Acott PD, Lee SHS, Bitter-Suermann H, Lawen JG, Crocker JFS. Infection concomitant with pediatric renal allograft rejection. *Transplantation.* 1996;62(5):689–91.
26. Shahinian VB, Muirhead N, Jevnikar AM, Leckie SH, Khakhar AK, Luke PP, et al. Epstein-Barr virus seronegativity is a risk factor for late-onset posttransplant lymphoproliferative

- disorder in adult renal allograft recipients. *Transplantation*. 2003;75(6):851–6.
27. Jamalidoust M, Namayandeh M, Pouladfar G, Ziyaeyan M. Post-Liver Transplant Cytomegalovirus (CMV) Reactivation, Graft, and Patient Survival Rates in Iranian Population. *Jundishapur J Microbiol*. 2021;14(3).
28. Fernández-García OA, García-Juárez I, Belaunzarán-Zamudio PF, Vilatoba M, Wisniowski-Yáñez A, Salomón-Ávila J, et al. Incidence of Cytomegalovirus disease and viral replication kinetics in seropositive liver transplant recipients managed under preemptive therapy in a tertiary-care center in Mexico City: a retrospective cohort study. *BMC Infect Dis*. 2022;22(1):1–7.
29. Varghese J, Subramanian S, Reddy MS, Shanmugam N, Balajee G, Srinivasan V, et al. Seroprevalence of cytomegalovirus in donors & opportunistic viral infections in liver transplant recipients. *Indian J Med Res*. 2017;145(4):558–60.
30. Dehghani M, kasiri K, mohamadi J, hasan pour K. Prevalence of Cytomegalovirus Infection in Candidates for Pediatric Liver Transplantation at Namazi Hospital of Shiraz During 2006-2009. *sjimu*. 2013;21(6):125-133.
31. Lizaola-Mayo BC, Rodriguez EA. Cytomegalovirus infection after liver transplantation. *World J Transplant*. 2020;10(7):183.
32. Gane E, Saliba F, Valdecasas GJC, O’Grady J, Pescovitz MD, Lyman S, et al. Randomised trial of efficacy and safety of oral ganciclovir in the prevention of cytomegalovirus disease in liver-transplant recipients. *Lancet*. 1997;350(9093):1729–33.
33. Abdullatif H, Dhawan A, Verma A. Epidemiology and Risk Factors for Viral Infections in Pediatric Liver Transplant Recipients and Impact on Outcome. *Viruses*. 2023;15(5):1059.
34. Halliday N, Smith C, Atkinson C, O’Beirne J, Patch D, Burroughs AK, et al. Characteristics of Epstein–Barr viraemia in adult liver transplant patients: A retrospective cohort study. *Transpl Int*. 2014;27(8):838–46.
35. KH M, SH Salari L. Prevalence of Epstein BARR virus infection and effecting factors in renal allograft recipients for controlling PTLD in imam khomeini hospital from 2001 to 2004. *J Shaheed Sadoughi Univ Med Sci Heal Serv*. 2007;15(3):53–60.
36. Rieder FJJ, Gröschel C, Kastner M-T, Kosulin K, Laengle J, Zadnikar R, et al. Human cytomegalovirus infection downregulates vitamin-D receptor in mammalian cells. *J Steroid Biochem Mol Biol*. 2017;165:356–62.
37. Beanrde A, Van Winden K, Frederick T, Kono N, Operskalski E, Pandian R, et al. Low maternal vitamin D is associated with increased risk of congenital and peri/postnatal transmission of Cytomegalovirus in women with HIV. *PLoS One*. 2020;15(2):e0228900.
38. Shirafkan H, Yazdani Charati J, Mozaffarpur SA, Khafri S, Akbari R, Oliaei F. Evaluation of Influential Factors in the Incidence Period of Cytomegalovirus after Renal Transplantation. *J Babol Univ Med Sci*. 2016;18(4):41–7.