

# An overview of nanoparticle production from plant gums and their action as antimicrobial agents

Melika Masoudi<sup>1</sup>, Amirhossein Tashakor<sup>1</sup>, Reza Abniki<sup>1</sup>, Davood Mansury<sup>1,2\*</sup>

<sup>1</sup>Department of Microbiology, School of Medicine, Isfahan University of Medical Sciences, Isfahan, Iran

<sup>2</sup>Nosocomial Infection Research Center, Isfahan University of Medical Sciences, Isfahan, Iran

## Abstract

There have been various chemicals and routines for the treatment of infections. The extensive use of antibiotics has led to serious issues including antibiotic resistance and serious side effects. The use of plant gum nanoparticles (NPs) is one of the several ways that can be employed greatly for the treatment of infections and have gained so much popularity by the scientists recently due to their several advantages over chemicals including being nontoxic and providing better tolerance to the patient. Several studies have been performed recently, stressing the undeniable advantages of these substances in the treatment of illnesses compared with their chemical counterparts. There are studies suggesting that these NPs have great potential in the treatment of multi-drug-resistant bacteria and that these substances have great anti-cancer effects due to their anti-inflammatory roles. Among various plant gums, Gum Arabia, gum Karaya, Kondagogu gum, and gum Tragacanth, Guar gum, and gum Ghatti have gathered more interest as anti-inflammatory subjects for studies because of their several pros including having more tissue bio-availability, being easy to use, etc. The use of plant gums can be limited due to a series of disadvantages but this can be untangled by using natural nanoparticles which can be synthesized via several ways including ultrasonic irradiation, etc. Among various metallic NPs, the most frequent of them in these studies are Silver nanoparticles (AgNPs) and Gold nanoparticles (AuNPs). According to these studies, AgNPs have a more bactericidal effect than AuNPs which is due to them being more of an antioxidant.

**Keywords:** Gum, Nanoparticles, Antimicrobial Activity, Treatment Agents

## 1. Introduction

The treatment of infections has always been considered as a significant factor. Thus, there has been various chemicals and routines for treatment of infections. The extensive use of antibiotics has led to serious issues including antibiotic resistance [1, 2]. There are also multi-drug resistant bacteria such as methicillin-resistant *Staphylococcus aureus* (MRSA), vancomycin-resistant Enterococci (VRE), multidrug-resistant Gram-negative bacteria (MDR-GNB) which

have led to noticeable problems in the world of medicine field [3-7]. There are also several microorganisms that can cause systematic illnesses which multi-drug therapy is used regularly for their treatment since most chemical drugs are designed to act against individual molecular targets. This itself can cause serious side effects [8, 9].

The use of plant gum nanoparticles (NPs) is one of the several ways that is employed greatly for the treatment of infections and this has been stated in

### \*Corresponding author:

Davood Mansury, PhD  
Department of Microbiology, School of Medicine,  
Isfahan University of Medical Sciences, Isfahan, Iran  
Isfahan, Iran, P.O. Box: 81746 -73461  
Tel/Fax: +98 31 37929098; +98 31 36688597  
Email: mansuryd@med.mui.ac.ir  
<http://orcid.org/0000-0003-0664-2007>

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various articles [8, 10-13]. The main source for plant gums are the gum producing trees, which they tend to grow inside the country's forests, and they can prepare various available materials. Plant-derived gums are consisted of polysaccharides and a few of them are applied medicinally for several years, including gum Tragacanth which has been used since 3rd century BC [14]. Various studies have shown the advantage of using green chemistry-based NPs for various purposes compared with using synthetic and chemical substances [15, 16]. Synthetic polymers have disadvantages, such as poor adaptation to the patient's body, high cost, and may also cause acute and chronic side effects. For example, polymethyl methacrylate (PMMA) can cause skin and eye irritation. Further disadvantages of these synthetic polymers are low biocompatibility, release of acidic products during degradation that may cause systemic or even local reactions, and prompt loss of mechanical durability [17, 18].

On the other hand, plant gums can have advantages in the pharmaceutical industry including being biodegradable, biocompatible, nontoxic, providing better tolerance to the patient and having fewer side effects [19]. They also do not cause allergies in humans, do not irritate the skin or eyes, and have low production costs [20]. There are articles showing that these plant gum nanoparticles were successful in the treatment of multi-drug resistant bacteria including MRSA [8]. Natural gums constitute a structurally diverse class of biological macromolecules with a broad range of physicochemical properties, therefore they can be loaded with various drugs and can have multi-target therapeutic effect. In this case, there won't be the need for consuming several drugs for the treatment of systemic illnesses [8].

That being said, the utilizing plant gums has its own series drawbacks. There is a high chance of bacterial contamination there is a high percentage of moisture in their components. They may also have a decrease in viscosity if they are kept in storages because of their exposer with water [18]. There are several ways in order to reduce these disadvantages and the most pragmatic way that has been used in various researches is turning plant gums into the nanoparticles. NPs are successfully applied in various fields, including pharmaceuticals and regenerative medicines and also the main reason for that, is related to having much better qualities including mechanical,

optical, thermal, biological, physical, and chemical aspects as compared to standard components [21]. They are also being used for measuring cellular organelles like liposome [22]. As a general definition, nanostructures are 1–100 nanometers in size during which case they need a high surface area-to-volume ratios and their reactivity is effected mostly by their difference in shape like spherical, conical, spiral, cylindrical, tubular, and hollow [21]. Green chemistry-based NPs are often being used for manufacturing products by adding durable components which can eliminate or decrease the existence and formation of toxic materials [10]. NPs created from plant gums may represent a nontoxic, effective and highly economical bio-resource for future medications [15]. Natural nanoparticles, improve the stability and bioavailability, as well as the biological distribution of natural products. They also significantly reduce the adverse effects of drug uptake. Therefore, gum based nano formulations are becoming much more popular [23].

Gum Arabic (GA), gum Karaya (GK), gum Tragacanth (GT), Kondagogu gum (KG), gum Ghatti (GG), and gum Guar are considered as some of the most popular plant gums that are being used regularly. So many researches has been recorded on tree gum polysaccharides, including their availability, chemical structures, and food or even medical applications. There also are several ways in order to create NPs from plant gums including mixing and agitation in a controlled environment, microwave (MW)-assisted technique, ultrasonic irradiation, etc. The biosynthesis of nanoparticles, nanofibers, and composites for supported tree gums would be very beneficial within the pursuit of relevance to medication for various health issues [24].

The purpose of this article is to review the beneficial medical aspects of these plant gum based NPs. There are several researches that have been done in order to show the advantages of these substances in infection and illness treatment. Various plant gums and routines have been used in order to create NPs with minimum side effects which will be discussed in this article.

## 2. Chemical property and chemical composition of plant gums

The plant gum exudates have a heterogeneous and sophisticated polysaccharides/carbohydrate

polymer [25]. The dry weight of the gum exudates is consisted of 2.45% of proteins, 0.85% of fats and 92.36% of carbohydrates. Other materials are the following: arabinose, xylitol, galactose and uremic acid (46.8: 10.9: 35.5: 6.0 mass ratio, respectively) with pieces of rhamnose, mannose and glucose. Also, gum exudates are full of minerals, like sodium, potassium, magnesium, calcium and iron [26]. The physical and functional properties of plant-based gums rely on their chemical compositions and molecular structures [2]. The chemical composition of plant polysaccharides is derived either from the identical sugar monomers (cellulose and starch), two various monomer units (alginate and hyaluronan) or various monosaccharide (galactose, arabinose, rhamnose and uremic acid) like gum acacia [27].

### 3. Green chemistry and nanomaterials

For the generation and stability of NPs, a variety of natural compounds can act as reducing and coating mediators [14]. Green chemistry refers to a set of concepts or practices that promote the development of goods and processes that decrease or eliminate the usage and creation of hazardous compounds. In the creation of NPs, current green nanotechnology methods frequently include the use of natural sources, non-hazardous solvents, biodegradable and biocompatible materials, and energy-efficient procedures. Biopolymers including cellulose, chitosan, dextran, and tree gums, for example, are frequently utilized as reducing and stabilizing agents in metal NP production. Plant-based ingredients (extracts, stems, gums, seeds, and fruits), among other biological sources, have been shown to be an efficient constituent for synthesizing nanoparticles while maintaining other important factors such as material cost, large-scale production capacity, and potential uses in a variety of applications. The pressure, temperature, solvent, and pH of the medium all play a role in the plant-based biogenic production of nanomaterials [28].

### 4. The green synthesis of nanoparticles and antibacterial applications

These natural gums are hydrophobic substances mostly obtained from plants or bacteria. Because the gum molecules are biological, they have a wide range of linear chain length, branching features, molecular weight, and other characteristics [29].

Various techniques can be used in order to create green nanoparticles by utilizing plant extracts for instance: (a) mixing and agitation in a controlled environment, (b) controlled heating in a certain temperature and pressure, (c) autoclaving, (d) microwave (MW)-assisted technique, (e) ultrasonic irradiation, and (f) UV/visible light irradiation. The use of ecologically benign solvent media (water and ionic, liquid-based green solvents) for the production of NPs shows that tree gums follow the cardinal principles of green chemistry. The presence of several functional groups (e.g.,  $-OH$ ,  $-COOH$ ,  $-CO$ , and  $CH_3CO-$ ) in the gum's structure, turns them into reducing agents. Natural gum fibers produce and stabilize NPs that are non-toxic to cells, making them ideal for a variety of functions such as medication administration, molecular imaging, and biomedical diagnostics. The usage of metal NPs (such as Ag, Au, Cu, CuO, and  $Fe_3O_4$ , among others) as coating materials in biomedical equipment requires both biocompatibility and antibacterial activity [24].

Plant gums such as gum Arabic, gum Karaya, gum Kondagogu, gum Tragacanth, gum Ghatti, Guar gum, Cashew gum, Gellan gum, Xanthan gum and Gum olibanu have been used as stabilizers and reducing agents in the production of metal/ metal oxide nanoparticles. Numerous studies have been conducted in this regard, which are briefly showed in Table 1 [10, 30-89].

Different studies have illustrated the benefits of using plant gum NPs as antibacterial agents which will be discussed in Table 2. There are several local (Iranian) plant gums that can be used as nanoparticles which can have great effect as antibacterial agents. These plant gums are including gum Arabia, *Prunus armeniaca* (Apricot), Neem gum, gum Kondagogu, gum Karaya, etc. [10]. Some of these plant gums and the bacteria that they have effect on has stated in Table 2. [8, 11, 61, 67, 90-94].

Table 1 Studies showed the green synthesis of NPs using plant gums.

Plant gums	Nanoparticle	Reference
Gum Arabic	Au, Zn, Magnetite, Cu, Ag, Se, Zein-curcumin, Chitosan/GA, Fe <sub>3</sub> O <sub>4</sub>	[30-47]
Gum Karaya	Ag, Cu, Au, Magnetite, Pt, Fe <sub>3</sub> O <sub>4</sub>	[31, 48-52]
Gum Kondagogu	Ag, Au, Cu, Pd, Ti, Pt, Fe <sub>3</sub> O <sub>4</sub> , Ag <sub>2</sub> S	[53-63]
Gum Tragacanth	Ag, ZnO, TiO <sub>2</sub> , Carbon dots, Au	[64-71]
Gum Ghatti	Pd, Magnetite, Ag, Au	[10, 72-74]
Guar gum	Ag, Au, Pt, Pd, Magnetite, Zn, Palmshell extract/ chitosan	[75-82]
Cashew gum	Ag, ZnO	[83, 84]
Gellan gum	TiO <sub>2</sub> , Ag	[85, 86]
Xanthan gum	Au	[87, 88]
Gum Olibanu	Ag	[89]

Table 2. Studies showed antibacterial effects of plant gums

Plant based NP	Preparation and essay	Results	Antibacterial activity	Reference
<i>Prunus domestica</i> gum loaded silver NPs	Disc diffusion method was used for antibacterial assay with using Gram-positive ( <i>S. aureus</i> ), Gram-negative ( <i>E. coli</i> ) and <i>P. aeruginosa</i> , three independent experiments were carried out for each bacterial strain with streptomycin as the positive control. Au/Ag-NPs (5 µg) were dissolved in DMSO and incubated at 30 °C for 24 h.	<i>P. domestica</i> gum-loaded silver nanoparticles can have potential antibacterial effect against <i>S. aureus</i> (19.7 ± 0.4 mm) and <i>E. coli</i> (14.4 ± 0.7 mm), and <i>P. aeruginosa</i> (13.1 ± 0.2 mm). Although this study suggests that streptomycin has an antibacterial effect of higher magnitude as compared to <i>P. domestica</i> gum-loaded silver nanoparticles against the tested bacterial strains (23.6 ± 0.8 mm, 21.8 ± 0.2 mm and 18.6 ± 0.3 mm).	Gram-positive ( <i>S. aureus</i> ), Gram-negative ( <i>E. coli</i> and <i>P. aeruginosa</i> )	[8]
<i>P. domestica</i> gum loaded gold NPs	Preparation and assessment performed like <i>P. domestica</i> gum loaded silver NPs.	Gum loaded gold nanoparticles had the least effect on foregoing bacteria ( <i>S. aureus</i> (10.5 ± 0.6 mm), <i>E. coli</i> (10 ± 0.4) mm and <i>P. aeruginosa</i> (8.2 ± 0.3 mm)) compared to <i>P. domestica</i> gum loaded silver NPs and streptomycin.	Gram-positive ( <i>S. aureus</i> ), Gram-negative ( <i>E. coli</i> and <i>P. aeruginosa</i> )	[8]

Plant based NP	Preparation and essay	Results	Antibacterial activity	Reference
GA-AgNPs (Gum acacia silver NPs) loaded with NP structures of HDN (fruit flavonoid)	Bactericidal assay was performed by incubating $10^8$ colony-forming units per mL of foregoing bacteria with various concentrations of GA-AgNPs-HDN and respective controls. For negative controls untreated bacterial culture were incubated with phosphate buffer saline (PBS), while 100 $\mu\text{g/mL}$ gentamicin treated bacteria were used as positive control.	Bactericidal effect of this nanoparticle was more significant on <i>Escherichia coli</i> K1 infections than MRSA infections, indicating that this component is more effective on Gram negative bacteria than Gram positive but overall these NPs have more bactericidal effects than chemicals.	Multi-drug resistant bacteria MRSA and Uropathogenic <i>E. coli</i> K1	[11]
Gum Kondagogu Copper nanoparticles (CuNPs)	The synthesized CuNPs are characterized by using Transmission Electron Microscopy (TEM), Scanning Electron Microscopy (SEM), UV visible spectroscopy, X-Ray Diffraction (XRD), FT-Raman spectroscopy and Fourier Transform Infrared Spectroscopy (FTIR) experimental methods.	Anti-biofilm effect of gum Kondagogu extract stabilized copper NPs against clinical isolate <i>Klebsiella pneumoniae</i> was demonstrated in this study.	<i>K. pneumoniae</i>	[61]
Gum Tragacanth (Astragalus gummifer) Silver nanoparticles	The well-diffusion method was used to study the antibacterial activity of the synthesized silver nanoparticles. Mueller Hinton agar plates were inoculated with 0.5 McFarland standard bacterial suspension, and 5 $\mu\text{g}$ of silver nanoparticles were added to the center well with a diameter of 6 mm. The nanoparticles used here were prepared with 0.1% gum solution containing $\text{AgNO}_3$ . Culture plates loaded with discs of antibiotic, erythromycin (15 $\mu\text{g}/\text{disc}$ ) were included as positive controls.	The inhibition zone of around $11.5 \pm 0$ mm was observed for the Gram-positive bacteria. For Gram-negative bacterial strains <i>E. coli</i> and <i>Pseudomonas aeruginosa</i> the inhibition zone was reported $9.5 \pm 0.4$ and $10.5 \pm 0$ respectively. Whereas, the negative control plates loaded with autoclaved gum did not produce any inhibition zone. In the case of positive control plates loaded with erythromycin discs, growth inhibition was noted.	Gram-positive bacterial strain <i>Staphylococcus aureus</i> and Gram-negative bacterial strains <i>E. coli</i> and <i>P. aeruginosa</i>	[67]
Chitosan/poly (vinyl alcohol)/guar gum (CS/PVA/GG)	After the preparation of a mixture of chitosan/poly (vinyl alcohol)/guar gum (CS/PVA/GG), the ratio of swelling, together with antimicrobial properties, was studied. These components were characterized by SEM, FTIR, and XRD.	SEM results showed that surface morphology was more affected by mixing and bonding ratios. Also, the FTIR and XRD confirmed the strong intermolecular bonding between polymers. The study suggests that these blends have great potential to be used against <i>Pasteurella multocida</i> , <i>S. aureus</i> , <i>E. coli</i> , and <i>Bacillus subtilis</i> .	<i>P. multocida</i> , <i>S. aureus</i> , <i>E. coli</i> , and <i>B. subtilis</i>	[90]

Plant based NP	Preparation and essay	Results	Antibacterial activity	Reference
Gum Karaya copper oxide (CuO) nanoparticles	The CuO nanoparticles were synthesized by a colloid-thermal synthesis process. The synthesized CuO was purified and dried to obtain different sizes of the CuO nanoparticles. The well diffusion method was used to study the antibacterial activity of the synthesized CuO nanoparticles. The zone of inhibition, minimum inhibitory concentration, and minimum bactericidal concentration were determined by the broth micro-dilution method.	The study suggests that gum Karaya copper oxide (CuO) nanoparticles can have significant bactericidal effect on both Gram-negative and positive cultures, specially smaller NPs ( $4.8 \pm 1.6$ nm) which are highly stable and have maximum zone of inhibition compared to the larger size of synthesized CuO nanoparticles ( $7.8 \pm 2.3$ nm).	Gram-negative and positive cultures	[91]
Kondagogu gum Gold nanoparticles (AuNPs)	After the preparation of these NPs, their concentration, and reaction time on the synthesis of AuNPs were investigated by using techniques like UV – visible spectroscopy, FTIR, DLS, TEM, and powder XRD.	The AuNPs showed good antibacterial activity against <i>E. coli</i> and <i>B. subtilis</i> .	<i>E. coli</i> and <i>B. subtilis</i>	[92]
Kondagogu Silver nanoparticles (AgNPs)	Variety of susceptibility assays were done in this study in order to demonstrate the antibacterial effects including: microbroth dilution, anti-biofilm activity, growth kinetics, cytoplasmic content leakage, membrane permeabilization, etc. The production of reactive oxygen species (ROS) and cell surface damage during bacterial nanoparticle interaction were also demonstrated using dichlorodihydrofluorescein diacetate, N-acetyl cysteine; and scanning electron microscopy and energy dispersive X-ray spectra.	The MIC values were lower by 3.2- and 16-folds for Gram-positive <i>S. aureus</i> and Gram-negative <i>E. coli</i> strains, respectively. The MBC values were lower by 4 and 50-folds. Thus, the biogenic AgNPs were found to be more potent bactericidal agents in terms of concentration. Results implies that this NPs has strong effects on biofilms, indicating that it can have great effect on drug resistant bacterial infections caused by biofilms. Also the growth curve stated a faster inhibition in Gram-negative bacteria as compared to Gram-positive.	Gram-positive <i>S. aureus</i> and Gram-negative <i>E. coli</i>	[93]

Plant based NP	Preparation and essay	Results	Antibacterial activity	Reference
Gum kondagogu Selenium nanoparticles (SeNPs)	Role of gum on synthesis and mean particle size was studied using ultraviolet-visible spectroscopy and dynamic light scattering. Size of the NPs were determined (from 44.4 to 200 nm) and mean particle size was 105.6 nm. Antibacterial potential of NPs was checked with well diffusion assay.	In this study, NPs exhibited growth inhibition activity against Gram-positive bacteria only. <i>B. subtilis</i> and <i>Micrococcus luteus</i> showed respective inhibition zones of 6.3 and 8.6 mm at 12 µg. this study implies that the tree gum stabilised Se NPs has more applicability as a potent antioxidant nutrition supplement at a much lower dose, in comparison with ionic Se.	<i>B. subtilis</i> and <i>M. luteus</i>	[94]

## 5. Conclusions and future prospects

Plant based synthesis and stabilization of metal/metal oxide NPs have been successfully implemented by many researchers worldwide. These techniques have various advantages including being more affordable physically and financially, having better drug distribution and having easier production. Based on these articles, using plant gums alone is less effective than their NP counterparts and in some occasions, they even may have side effects on human's body [95].

Various metallic or non-metallic NPs can be created and added to these plant gums. Most frequent of them in these studies are silver nanoparticles (AgNPs) and gold nanoparticles (AuNPs). AuNPs have exceptional stability against oxidation and therefore they may play a significant role in the advancement of clinically useful diagnostic and therapeutic nanomedicines. However, according to various studies, they are less effective against microorganisms than AgNPs. AgNPs have a strong bactericidal and catalytic effect according to various studies, they are extremely beneficial for preventing drug resistant bacteria which will be a huge issue in the future [96-98].

The influence of different parameters such as gum particle size, concentration of gum, concentration of silver nitrate and reaction time on the synthesis of nanoparticles is quite significant in various studies [11, 99, 100]. The influence of the nanoparticles size on their effectiveness is well defined for a NP like copper oxide which smaller NPs can have more bactericidal effects compared to their bigger counter parts [101].

Thus using the right concentration and technique for making these NPs is very important and it should be considered.

The future use of tree gums also relies on the development of ultralight weight, high, strength, bio-based, biodegradable, porous, etc. Each year scientists are getting keener on researching about these green NPs because of various reasons including the significant growth in the number of antibiotic resistant bacteria or climate change.

### Authors' contributions

Study design, supervision, conceptualization, and critical revisions: DM. Searching, data collection and drafting: MM, AT. The final manuscript has been read and approved for submission by all the named authors.

### Conflict of interests

The authors reported no potential conflict of interest.

### Ethical declarations

None.

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## References

- Ventola CLJP, therapeutics. The antibiotic resistance crisis: part 1: causes and threats. 2015; 40(4):277-83.
- Mansury D, Motamedifar M, Sarvari J, Shirazi B, Khaledi A. Antibiotic susceptibility pattern and identification of extended

- spectrum  $\beta$ -lactamases (ESBLs) in clinical isolates of *Klebsiella pneumoniae* from Shiraz, Iran. *Iran J Microbiol.* 2016; 8(1):55-61.
3. Siddiqui AH, Koirala J. Methicillin Resistant *Staphylococcus aureus*. *StatPearls* [internet]. 2020.
4. Motamedifar M, Sarai HSE, Mansury D. Patterns of constitutive and inducible clindamycin resistance in *Staphylococcus aureus* isolated from clinical samples by D-test method, shiraz, southwest of Iran. *Galen Med J.* 2014; 3(4):216-21.
5. Levitus M, Rewane A, Perera TB. Vancomycin-Resistant *Enterococci*. 2023. In: *StatPearls* [Internet].
6. Bassetti M, Peghin M, Vena A, Giacobbe DRJFim. Treatment of infections due to MDR Gram-negative bacteria. *Front Med (Lausanne).* 2019; 6:74.
7. Tanhaeian A, Damavandi MS, Mansury D, Ghaznini K. Expression in eukaryotic cells and purification of synthetic gene encoding enterocin P: a bacteriocin with broad antimicrobial spectrum. *AMB Express.* 2019; 9(1):6.
8. Islam NU, Amin R, Shahid M, Amin M, Zaib S, Iqbal J. A multi-target therapeutic potential of *Prunus domestica* gum stabilized nanoparticles exhibited prospective anticancer, antibacterial, urease-inhibition, anti-inflammatory and analgesic properties. *BMC Complement Altern Med.* 2017; 17(1):1-17.
9. Deps PD, Nasser S, Guerra P, Simon M, Birshner RDC, Rodrigues LC. Adverse effects from multi-drug therapy in leprosy: a Brazilian study. *Lepr Rev.* 2007; 78(3):216-22.
10. Irvani S. Plant gums for sustainable and eco-friendly synthesis of nanoparticles: recent advances. *Inorg Nano-Met Chem.* 2020; 50(6):469-88.
11. Anwar A, Masri A, Rao K, Rajendran K, Khan NA, Shah MR, et al. Antimicrobial activities of green synthesized gums-stabilized nanoparticles loaded with flavonoids. *Sci Rep.* 2019; 9(1):1-12.
12. Sharma G, Kalra SK, Tejan N, Ghoshal U. Nanoparticles based therapeutic efficacy against *Acanthamoeba*: Updates and future prospect. *Exp Parasitol.* 2020; 218:108008.
13. Escárcega-González CE, Garza-Cervantes JA, Vazquez-Rodríguez A, Montelongo-Peralta LZ, Treviño-Gonzalez MT, Castro EDB, et al. In vivo antimicrobial activity of silver nanoparticles produced via a green chemistry synthesis using *Acacia rigidula* as a reducing and capping agent. *Int J Nanomed.* 2018; 13:2349.
14. Huang J, Lin L, Sun D, Chen H, Yang D, Li Q. Bio-inspired synthesis of metal nanomaterials and applications. *Chem Soc Rev.* 2015; 44(17):6330-74.
15. Darroudi M, Yazdi MET, Amiri MS. Plant-mediated biosynthesis of nanoparticles. 21st century nanoscience—A handbook: CRC Press; 2020. p. 1—18.
16. Hashemzadeh MR, Yazdi MET, Amiri MS, Mousavi SH. Stem cell therapy in the heart: Biomaterials as a key route. *Tissue Cell.* 2021:101504.
17. Khan I, Saeed K, Khan I. Nanoparticles: Properties, applications and toxicities. *Arab J Chem.* 2019; 12(7):908-31.
18. Amiri MS, Mohammadzadeh V, Yazdi MET, Barani M, Rahdar A, Kyzas GZ. Plant-Based Gums and Mucilages Applications in Pharmacology and Nanomedicine: A Review. *Molecules.* 2021; 26(6):1770.
19. Bhosale RR, Osmani RAM, Moin A. Natural gums and mucilages: a review on multifaceted excipients in pharmaceutical science and research. *Int J Pharmacogn Phytochem Res.* 2014; 15(4):901-12.
20. Abulafatih H. Medicinal plants in southwestern Saudi Arabia. *Econ Bot.* 1987; 41(3):354-60.
21. Rastogi A, Zivcak M, Sytar O, Kalaji HM, He X, Mbarki S, et al. Impact of metal and metal oxide nanoparticles on plant: a critical review. *Front Chem.* 2017; 5:78.
22. Mansury D, Ghazvini K, Jamehdar SA, Badiie A, Tafaghodi M, Nikpoor AR, et al. Increasing cellular immune response in liposomal formulations of DOTAP encapsulated by fusion protein Hsp<sub>90</sub>, PPE44, and Esxv, as a potential tuberculosis vaccine candidate. *Rep Biochem Mol Bio.* 2019; 7(2):156.
23. Patra JK, Das G, Fraceto LF, Campos EVR, del Pilar Rodriguez-Torres M, Acosta-Torres LS, et al. Nano based drug delivery systems: recent developments and future prospects. *J Nanobiotechnology.* 2018; 16(1):1-33.
24. Padil VV, Zare EN, Makvandi P, Černík M. Nanoparticles and nanofibres based on tree gums: Biosynthesis and applications. *Compr Anal Chem.* 2021. p. 223-65.
25. Mirhosseini H, Amid BT. A review study on chemical composition and molecular structure of newly plant gum exudates and seed gums. *Food Res Int.* 2012; 46(1):387-98.
26. Mahfoudhi N, Chouaibi M, Donsi F, Ferrari G, Hamdi S. Chemical composition and functional properties of gum exudates from the trunk of the almond tree (*Prunus dulcis*). *Food Sci Technol Int.* 2012; 18(3):241-50.
27. Williams P. Introduction to food hydrocolloids. IN PHILLIPS, GO & WILLIAMS, PA (Eds) *Handbook of Hydrocolloids*, p 1-19. Woodhead publishing Limited; 2000.
28. Nguyen NH, Padil VVT, Slaveykova VI, Černík M, Ševců A. Green synthesis of metal and metal oxide nanoparticles and their effect on the unicellular alga *Chlamydomonas reinhardtii*. *Nanoscale Res Lett.* 2018; 13(1):1-13.
29. Sharma G, Sharma S, Kumar A, Ala'a H, Naushad M, Ghfar AA, et al. Guar gum and its composites as potential materials for diverse applications: A review. *Carbohydr Polym.* 2018; 199:534-45.
30. Kong H, Yang J, Zhang Y, Fang Y, Nishinari K, Phillips GO. Synthesis and antioxidant properties of gum arabic-stabilized selenium nanoparticles. *Int J Biol Macromol.* 2014; 65:155-62.
31. Kattumuri V, Katti K, Bhaskaran S, Boote EJ, Casteel SW, Fent GM, et al. Gum arabic as a phytochemical construct for the stabilization of gold nanoparticles: in vivo pharmacokinetics and X-ray-contrast-imaging studies. *Small.* 2007; 3(2):333-41.
32. Fent GM, Casteel SW, Kim DY, Kannan R, Katti K, Chanda N, et al. Biodistribution of maltose and gum arabic hybrid gold nanoparticles after intravenous injection in juvenile swine. *Nanomed: Nanotech, Biol Med.* 2009; 5(2):128-35.
33. Shalaby T, El-Dine RSS, El-Gaber S. Green Synthesis Of gold Nanoparticles Using Cumin Seeds and Gum Arabic: Studying Their Photothermal Efficiency. *Nanosci Nanotechnol.* 2015; 5(4):89-96.
34. Chopra M, Bernela M, Kaur P, Manuja A, Kumar B, Thakur R. Alginate/gum acacia bipolymeric nanohydrogels—Promising carrier for Zinc oxide nanoparticles. *Int J Biol Macromol.* 2015; 72:827-33.
35. Roque A, Wilson Jr O. Adsorption of gum Arabic on bioceramic nanoparticles. *Mater Sci Eng C.* 2008; 28(3):443-7.
36. Roque AC, Bicho A, Batalha IL, Cardoso AS, Hussain A. Biocompatible and bioactive gum Arabic coated iron oxide magnetic nanoparticles. *J Biotechnol.* 2009; 144(4):313-20.



37. Dong C, Cai H, Zhang X, Cao C. Synthesis and characterization of monodisperse copper nanoparticles using gum acacia. *Physica E Low Dimens Syst Nanostruct.* 2014; 57:12-20.
38. Mohan YM, Raju KM, Sambasivudu K, Singh S, Sreedhar B. Preparation of acacia-stabilized silver nanoparticles: A green approach. *J Appl Polym Sci.* 2007; 106(5):3375-81.
39. Akele ML, Assefa AG, Alle M. Microwave-assisted green synthesis of silver nanoparticles by using gum acacia: Synthesis, characterization and catalytic activity studies. *Int J Green Chem Bioprocess.* 2015; 5:21-7.
40. Dong C, Zhang X, Cai H, Cao C. Facile and one-step synthesis of monodisperse silver nanoparticles using gum acacia in aqueous solution. *J Mol Liq.* 2014; 196:135-41.
41. Djajadisatra J, Sutriyo PP, Purnamasari P, Pujiyanto A. Antioxidant activity of gold nanoparticles using gum arabic as a stabilizing agent. *Int J Pharm Pharm Sci.* 2014; 6(7):462-5.
42. Wu C-C, Chen D-H. Facile green synthesis of gold nanoparticles with gum arabic as a stabilizing agent and reducing agent. *Gold Bull.* 2010; 43(4):234-40.
43. Baruwati B, Varma RS. High value products from waste: grape pomace extract-a three-in-one package for the synthesis of metal nanoparticles. *Chem Sus Chem.* 2009; 2(11):1041-4.
44. Chockalingam AM, Babu HKRR, Chittor R, Tiwari JP. Gum arabic modified Fe<sub>3</sub>O<sub>4</sub> nanoparticles cross linked with collagen for isolation of bacteria. *J Nanobiotechnology.* 2010; 8(1):1-9.
45. Chawla P, Kumar N, Bains A, Dhull SB, Kumar M, Kaushik R, et al. Gum arabic capped copper nanoparticles: Synthesis, characterization, and applications. *Int J Biol Macromol.* 2020; 146:232-42.
46. Zamani H, Rastegari B, Varamini M. Antioxidant and anti-cancer activity of Dunaliella salina extract and oral drug delivery potential via nano-based formulations of gum Arabic coated magnetite nanoparticles. *J Drug Deliv Sci Technol.* 2019; 54:101278.
47. Chen G, Fu Y, Niu F, Zhang H, Li X, Li X. Evaluation of the colloidal/chemical performance of core-shell nanoparticle formed by zein and gum Arabic. *Colloids Surf A: Physicochem Eng Asp.* 2019; 560:130-5.
48. Sharkawy A, Barreiro MF, Rodrigues AE. Preparation of chitosan/gum Arabic nanoparticles and their use as novel stabilizers in oil/water Pickering emulsions. *Carbohydr Polym.* 2019; 224:115190.
49. Padil VVT, Černík M. Poly (vinyl alcohol)/gum karaya electrospun plasma treated membrane for the removal of nanoparticles (Au, Ag, Pt, CuO and Fe<sub>3</sub>O<sub>4</sub>) from aqueous solutions. *J Hazard Mater.* 2015; 287:102-10.
50. Pooja D, Panyaram S, Kulhari H, Reddy B, Rachamalla SS, Sistla R. Natural polysaccharide functionalized gold nanoparticles as biocompatible drug delivery carrier. *Int J Biol Macromol.* 2015; 80:48-56.
51. Padil VVT, Černík M. Green synthesis of copper oxide nanoparticles using gum karaya as a biotemplate and their antibacterial application. *Int J Nanomedicine.* 2013; 8:889.
52. Vellora Thekkae Padil V, Nguyen NH, Ševců A, Černík M. Fabrication, characterization, and antibacterial properties of electrospun membrane composed of gum karaya, polyvinyl alcohol, and silver nanoparticles. *J Nanomater.* 2015; 2015.
53. Gangapuram BR, Bandi R, Dadigala R, Kotu GM, Guttena V. Facile green synthesis of gold nanoparticles with carboxymethyl gum karaya, selective and sensitive colorimetric detection of copper (II) ions. *J Clust Sci.* 2017; 28(5):2873-90.
54. Vellora Thekkae Padil V, Rouha M, Černík M. Hydrocolloid-stabilized magnetite for efficient removal of radioactive phosphates. *Biomed Res Int.* 2014; 2014 :504760.
55. Venkatesham M, Ayodhya D, Madhusudhan A, Kumari AS, Veerabhadram G, Mangatayaru KG. A novel green synthesis of silver nanoparticles using gum karaya: characterization, antimicrobial and catalytic activity studies. *J Clust Sci.* 2014; 25(2):409-22.
56. Kora AJ, Sashidhar R, Arunachalam J. Gum kondagogu (*Cochlospermum gossypium*): a template for the green synthesis and stabilization of silver nanoparticles with antibacterial application. *Carbohydr Polym.* 2010; 82(3):670-9.
57. Saravanan P, Vinod V, Sreedhar B, Sashidhar R. Gum kondagogu modified magnetic nano-adsorbent: An efficient protocol for removal of various toxic metal ions. *Mater Sci Eng C.* 2012; 32(3):581-6.
58. Vinod V, Saravanan P, Sreedhar B, Devi DK, Sashidhar R. A facile synthesis and characterization of Ag, Au and Pt nanoparticles using a natural hydrocolloid gum kondagogu (*Cochlospermum gossypium*). *Colloids Surf B Biointerfaces.* 2011; 83(2):291-8.
59. Rastogi L, Sashidhar R, Karunasagar D, Arunachalam J. Gum kondagogu reduced/stabilized silver nanoparticles as direct colorimetric sensor for the sensitive detection of Hg<sup>2+</sup> in aqueous system. *Talanta.* 2014; 118:111-7.
60. Reddy GB, Madhusudhan A, Ramakrishna D, Ayodhya D, Venkatesham M, Veerabhadram G. Green chemistry approach for the synthesis of gold nanoparticles with gum kondagogu: characterization, catalytic and antibacterial activity. *J Nanostructure Chem.* 2015; 5(2):185-93.
61. Suresh Y, Annapurna S, Singh A, Chetana A, Pasha C, Bhikshamaiah G. Characterization and evaluation of anti-biofilm effect of green synthesized copper nanoparticles. *Mater Today Proc.* 2016; 3(6):1678-85.
62. Saranya KS, Vellora Thekkae Padil V, Senan C, Pilankatta R, Saranya K, George B, et al. Green synthesis of high temperature stable anatase titanium dioxide nanoparticles using Gum Kondagogu: Characterization and solar driven photocatalytic degradation of organic dye. *Nanomaterials.* 2018; 8(12):1002.
63. Padil VVT, Stuchlík M, Černík M. Plasma modified nanofibres based on gum kondagogu and their use for collection of nanoparticulate silver, gold and platinum. *Carbohydr Polym.* 2015; 121:468-76.
64. Rastogi L, Beedu SR, Kora AJ. Facile synthesis of palladium nanocatalyst using gum kondagogu (*Cochlospermum gossypium*): a natural biopolymer. *IET nanobiotechnol.* 2015; 9(6):362-7.
65. Venkateshaiah A, Silvestri D, Ramakrishnan RK, Waclawek S, Padil VV, Černík M, et al. Gum kondagogu/reduced graphene oxide framed platinum nanoparticles and their catalytic role. *Molecules.* 2019; 24(20):3643.
66. Ayodhya D, Veerabhadram G. Green synthesis, characterization, photocatalytic, fluorescence and antimicrobial activities of *Cochlospermum gossypium* capped Ag<sub>2</sub>S nanoparticles. *J Photochem Photobiol B.* 2016; 157:57-69.
67. Rao K, Imran M, Jabri T, Ali I, Perveen S, Ahmed S, et al. Gum tragacanth stabilized green gold nanoparticles as cargos for Naringin loading: A morphological investigation through AFM. *Carbohydr Polym.* 2017; 174:243-52.

68. Kora AJ, Arunachalam J. Green fabrication of silver nanoparticles by gum tragacanth (*Astragalus gummifer*): a dual functional reductant and stabilizer. *J Nanomater.* 2012; 2012.
69. Ghayempour S, Montazer M, Rad MM. Tragacanth gum biopolymer as reducing and stabilizing agent in biosynthesis of urchin-like ZnO nanorod arrays: A low cytotoxic photocatalyst with antibacterial and antifungal properties. *Carbohydr Polym.* 2016; 136:232-41.
70. Montazer M, Keshvari A, Kahali P. Tragacanth gum/nano silver hydrogel on cotton fabric: In-situ synthesis and antibacterial properties. *Carbohydr Polym.* 2016; 154:257-66.
71. Ranjbar-Mohammadi M, Rahimdokht M, Pajootan E. Low cost hydrogels based on gum Tragacanth and TiO<sub>2</sub> nanoparticles: characterization and RBFNN modelling of methylene blue dye removal. *Int J Biol Macromol.* 2019; 134:967-75.
72. Ghayempour S, Montazer M. Ultrasound irradiation based in-situ synthesis of star-like Tragacanth gum/zinc oxide nanoparticles on cotton fabric. *Ultrason Sonochem.* 2017; 34:458-65.
73. Darroudi M, Sabouri Z, Oskuee RK, Zak AK, Kargar H, Abd Hamid MHN. Sol-gel synthesis, characterization, and neurotoxicity effect of zinc oxide nanoparticles using gum tragacanth. *Ceram Int.* 2013; 39(8):9195-9.
74. Moradi S, Sadrjavadi K, Farhadian N, Hosseinzadeh L, Shahlai M. Easy synthesis, characterization and cell cytotoxicity of green nano carbon dots using hydrothermal carbonization of Gum Tragacanth and chitosan bio-polymers for bioimaging. *J Mol Liq.* 2018; 259:284-90.
75. Kora AJ, Beedu SR, Jayaraman A. Size-controlled green synthesis of silver nanoparticles mediated by gum ghatti (*Anogeissus latifolia*) and its biological activity. *Org Med Chem Lett.* 2012; 2(1):1-10.
76. Mittal H, Mishra SB. Gum ghatti and Fe<sub>3</sub>O<sub>4</sub> magnetic nanoparticles based nanocomposites for the effective adsorption of rhodamine B. *Carbohydrate polymers.* 2014; 101:1255-64.
77. Alam MS, Garg A, Pottou FH, Saifullah MK, Tareq AI, Manzoor O, et al. Gum ghatti mediated, one pot green synthesis of optimized gold nanoparticles: Investigation of process-variables impact using Box-Behnken based statistical design. *Int J Biol Macromol.* 2017; 104:758-67.
78. Pandey S, Mishra SB. Catalytic reduction of p-nitrophenol by using platinum nanoparticles stabilised by guar gum. *Carbohydr Polym.* 2014; 113:525-31.
79. Dhiman N, Singh A, Verma NK, Ajaria N, Patnaik S. Statistical optimization and artificial neural network modeling for acridine orange dye degradation using in-situ synthesized polymer capped ZnO nanoparticles. *J Colloid Interface Sci.* 2017; 493:295-306.
80. Vanamudan A, Sudhakar PP. Biopolymer capped silver nanoparticles with potential for multifaceted applications. *Int J Biol Macromol.* 2016; 86:262-8.
81. Pandey S, Goswami GK, Nanda KK. Green synthesis of polysaccharide/gold nanoparticle nanocomposite: an efficient ammonia sensor. *Carbohydr Polym.* 2013; 94(1):229-34.
82. Rastogi PK, Ganesan V, Krishnamoorthi S. Palladium nanoparticles incorporated polymer-silica nanocomposite based electrochemical sensing platform for nitrobenzene detection. *Electrochim Acta.* 2014; 147:442-50.
83. Murali R, Vidhya P, Thanikaivelan P. Thermoresponsive magnetic nanoparticle-aminated guar gum hydrogel system for sustained release of doxorubicin hydrochloride. *Carbohydr Polym.* 2014; 110:440-5.
84. Pandey S, Goswami GK, Nanda KK. Green synthesis of biopolymer-silver nanoparticle nanocomposite: An optical sensor for ammonia detection. *Int J Biol Macromol.* 2012; 51(4):583-9.
85. Vanaamudan A, Sadhu M, Pamidimukkala P. Chitosan-Guar gum blend silver nanoparticle bionanocomposite with potential for catalytic degradation of dyes and catalytic reduction of nitrophenol. *J Mol Liq.* 2018; 271:202-8.
86. Souza JMT, de Araujo AR, de Carvalho AMA, Amorim AdGN, Daboit TC, de Almeida JRdS, et al. Sustainably produced cashew gum-capped zinc oxide nanoparticles show antifungal activity against *Candida parapsilosis*. *J Clean Prod.* 2020; 247:119085.
87. Araruna FB, de Oliveira TM, Quelemes PV, de Araujo Nobre AR, Placido A, Vasconcelos AG, et al. Antibacterial application of natural and carboxymethylated cashew gum-based silver nanoparticles produced by microwave-assisted synthesis. *Carbohydr Polym.* 2020; 241:115260.
88. Ismail NA, Amin KAM, Majid FAA, Razali MH. Gellan gum incorporating titanium dioxide nanoparticles biofilm as wound dressing: Physicochemical, mechanical, antibacterial properties and wound healing studies. *Mater Sci Eng C Mater Biol Appl.* 2019; 103:109770.
89. Zhai X, Li Z, Shi J, Huang X, Sun Z, Zhang D, et al. A colorimetric hydrogen sulfide sensor based on gellan gum-silver nanoparticles bionanocomposite for monitoring of meat spoilage in intelligent packaging. *Food Chem.* 2019; 290:135-43.
90. Alle M, Kim TH, Park SH, Lee S-H, Kim J-C. Doxorubicin-carboxymethyl xanthan gum capped gold nanoparticles: microwave synthesis, characterization, and anti-cancer activity. *Carbohydr Polym.* 2020; 229:115511.
91. Singh J, Kumar S, Dhaliwal A. Controlled release of amoxicillin and antioxidant potential of gold nanoparticles-xanthan gum/poly (Acrylic acid) biodegradable nanocomposite. *J Drug Deliv Sci Technol.* 2020; 55:101384.
92. Kora AJ, Sashidhar R, Arunachalam J. Aqueous extract of gum olibanum (*Boswellia serrata*): a reductant and stabilizer for the biosynthesis of antibacterial silver nanoparticles. *Process Biochem.* 2012; 47(10):1516-20.
93. Iqbal DN, Shafiq S, Khan SM, Ibrahim SM, Abubshait SA, Nazir A, et al. Novel chitosan/guar gum/PVA hydrogel: Preparation, characterization and antimicrobial activity evaluation. *Int J Biol Macromol.* 2020; 164:499-509.
94. Seku K, Gangapuram BR, Pejjai B, Hussain M, Hussaini SS, Golla N, et al. Eco-friendly synthesis of gold nanoparticles using carboxymethylated gum *Cochlospermum gossypium* (CMGK) and their catalytic and antibacterial applications. *Chem Pap.* 2019; 73(7):1695-704.
95. Laubach J, Joseph M, Brenza T, Gadhamshetty V, Sani RKJJocr. Exopolysaccharide and biopolymer-derived films as tools for transdermal drug delivery. *J Control Release.* 2021; 329:971-987.
96. Shirzadi-Ahodashi M, Mizwari ZM, Hashemi Z, Rajabalipour S, Ghoreishi SM, Mortazavi-Derazkola S, et al. Discovery of high antibacterial and catalytic activities of biosynthesized silver nanoparticles using *C. fruticosus* (CF-AgNPs) against multi-drug resistant clinical strains and hazardous pollutants. *Environ Technol Innov.* 2021;23:101607.

97. Nouri A, Yaraki MT, Lajevardi A, Rezaei Z, Ghorbanpour M, Tanzifi MJC, et al. Ultrasonic-assisted green synthesis of silver nanoparticles using *Mentha aquatica* leaf extract for enhanced antibacterial properties and catalytic activity. *Colloids Interface Sci. Commun.* 2020;35:100252.
98. Cheng D, Zhang Y, Liu Y, Bai X, Ran J, Bi S, et al. Mussel-inspired synthesis of filter cotton-based AgNPs for oil/water separation, antibacterial and catalytic application. *Mater Today Commun.* 2020;25:101467.
99. Dong Y, Zhu H, Shen Y, Zhang W, Zhang LJPo. Antibacterial activity of silver nanoparticles of different particle size against *Vibrio Natriegens*. *PLoS One.* 2019; 14(9):e0222322.
100. Lallo da Silva B, Abuçafy MP, Berbel Manaia E, Oshiro Junior JA, Chiari-Andréo BG, Pietro RCR, et al. Relationship between structure and antimicrobial activity of zinc oxide nanoparticles: An overview. *Int J Nanomedicine.* 2019; 14:9395-9410.
101. Mohd Yusof H, Mohamad R, Zaidan UH, Abdul Rahman NAJJoas, biotechnology. Microbial synthesis of zinc oxide nanoparticles and their potential application as an antimicrobial agent and a feed supplement in animal industry: a review. *J Anim Sci Biotechnol.* 2019; 10:57.